

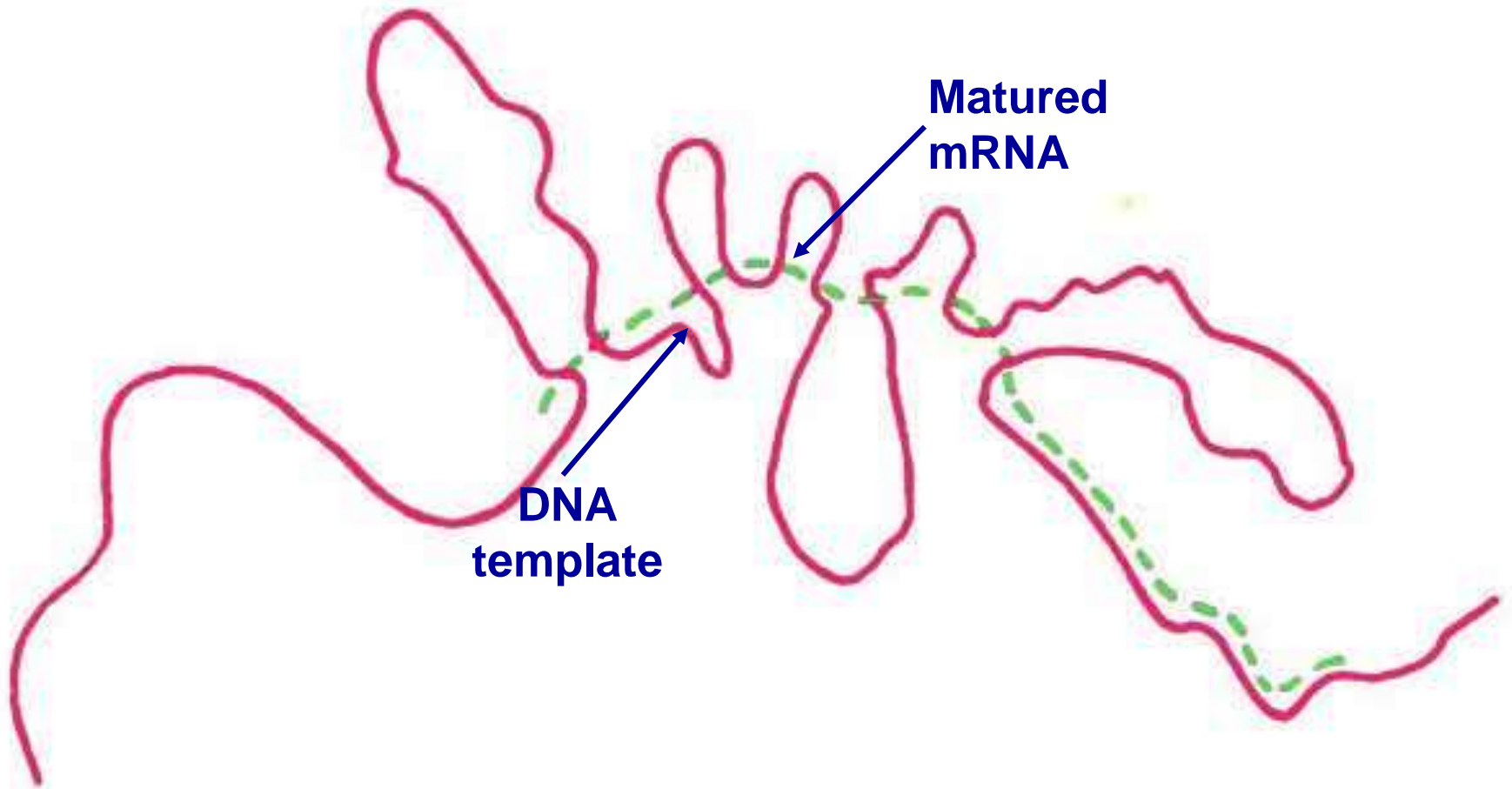
**SOS in Biochemistry, Jiwaji University, Gwalior**

**M.Sc. II Semester (2019-20)**

**Paper BCH 201: Fundamentals of Molecular Biology (Unit IV)**

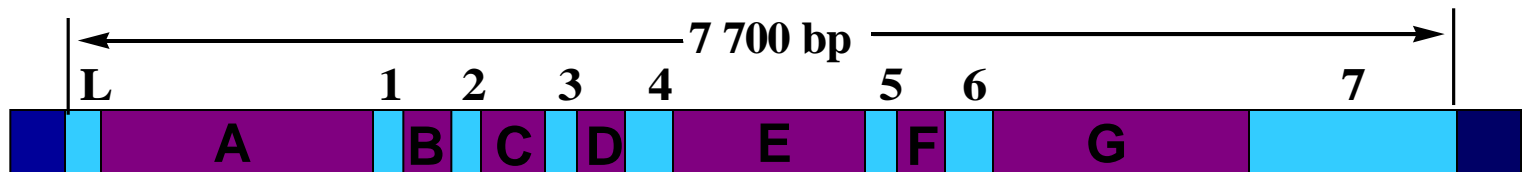
# **Nuclear pre-mRNA Splicing**

**The matured mRNAs are much shorter than the DNA templates.**



# Split Gene

The structural genes are composed of **coding** and **non-coding** regions that are alternatively separated.



**A~G** no-coding region    **1~7** coding region

**NOBEL PRIZE – 1993**

**PHYSIOLOGY OR MEDICINE**

**For the discovery of**

**Split Genes**

# **The Nobel Prize was shared by two American Scientists:**

- 1. Richard J. Roberts, New England Biolabs, Beverly, MA, USA (1/2)**
- 2. Phillip A. Sharp, Massachusetts Institute of Technology (MIT), Center for Cancer Research, Cambridge, MA, USA (1/2)**



**Richard J. Roberts**

**Born:** 6 September 1943, Derby, United Kingdom

**Field:** Genetics, Molecular Biology

**Prize share:** 1/2



**Phillip A. Sharp**

**Born:** 6 June 1944, Falmouth, KY, USA

**Field:** Genetics, Molecular Biology

**Prize share:** 1/2

**Work: Split Genes**

# Exon and Intron

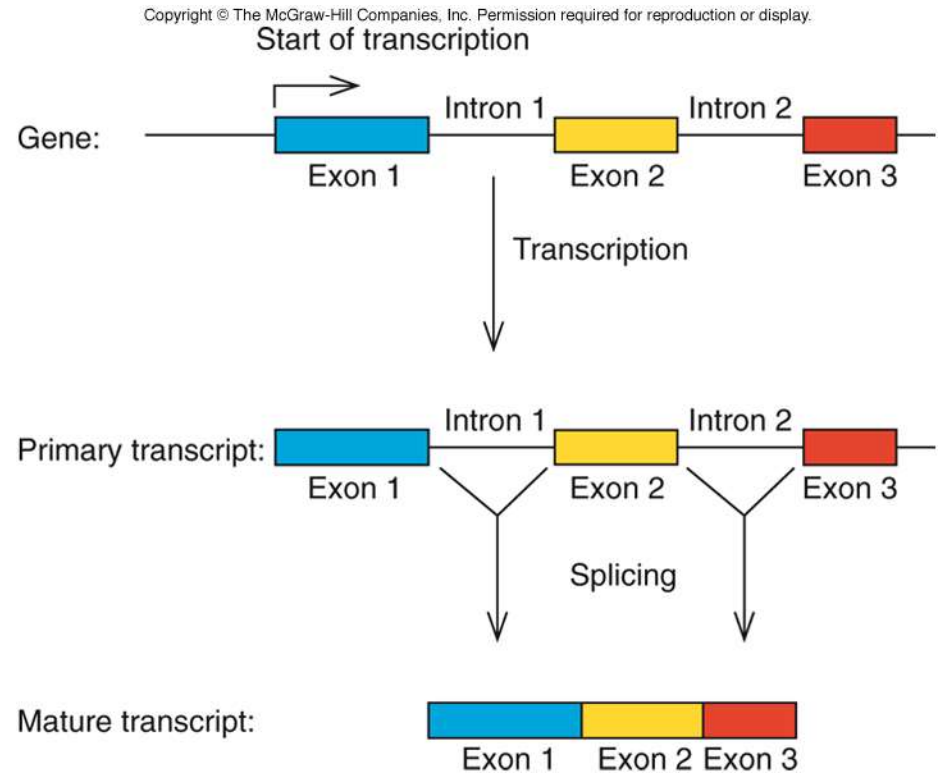
**Exons** are the **coding** sequences that appear on split genes and primary transcripts, and will be expressed to matured mRNA.

**Introns** are the **non-coding** sequences that are transcribed into primary mRNAs, and will be cleaved out in the later splicing process.



# Splicing Outline

- **Introns are transcribed along with exons in the primary transcript**
- **Introns are removed as the exons are spliced together**



# Types of Introns

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Intron Type	Where Found
GU-AG introns	Eukaryotic nuclear pre-mRNA
AU-AC introns	Eukaryotic nuclear pre-mRNA
Group I	Eukaryotic nuclear pre-mRNA, organelle RNAs, a few bacterial RNAs
Group II	Organelle RNAs, a few prokaryotic RNAs
Group III	Organelle RNAs
Twintrons (composites of two and/or more group II or III introns)	Organelle RNAs
Pre-tRNA introns	Eukaryotic nuclear pre-tRNAs
Archaeal introns	Various RNAs

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# TYPES OF INTRONS

Introns in all genes can be divided into three general classes:

(Except nuclear tRNA coding genes)

## 1. Nuclear pre-mRNA Introns

(With GU-----AG dinucleotide at 5' & 3' ends and a branch site near the 3' end)

## 2. Group I Introns

(Found in organelles & bacteria)

(Also found in nucleus of lower eukaryotes)

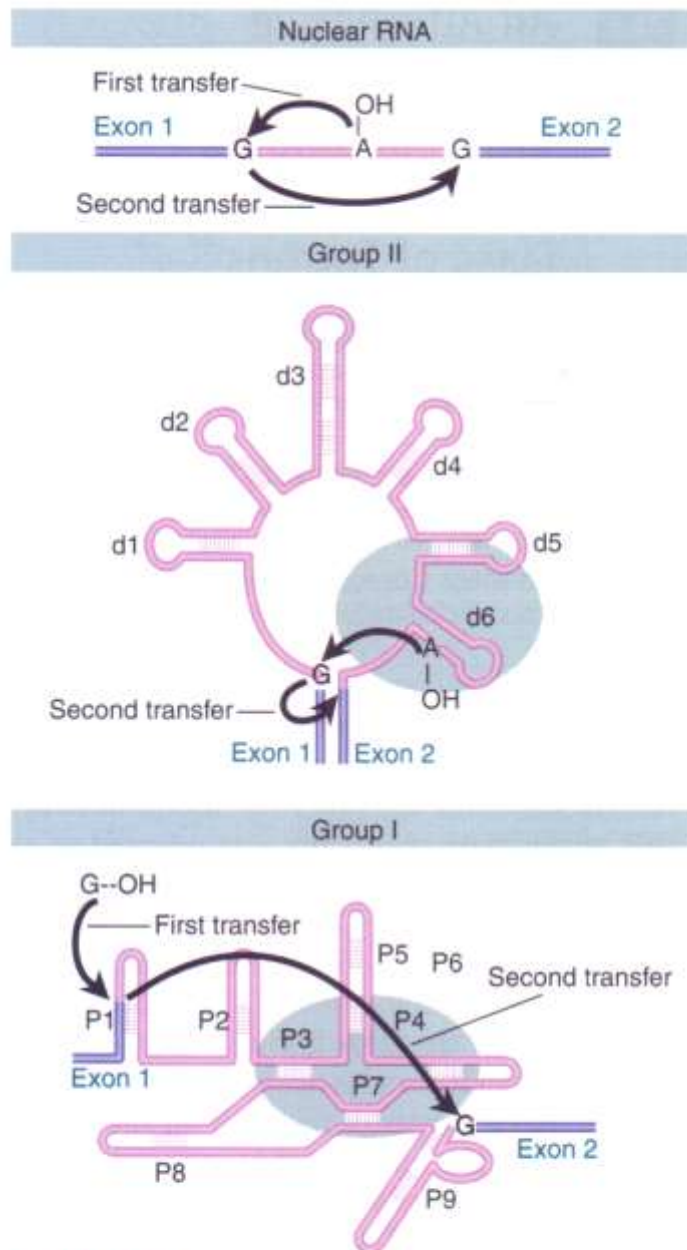
## 3. Group II Introns

(Found in organelles & bacteria)

...Gp I introns are more common than Gp II introns and both possess auto splicing / self splicing property

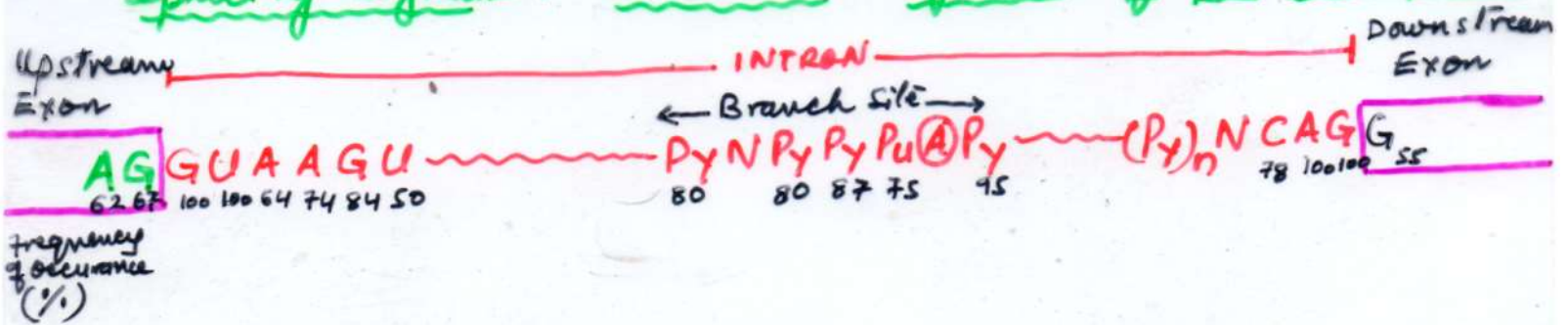
...Self splicing introns are classified according to their internal organization

(Each can be folded into a typical type of secondary structure)



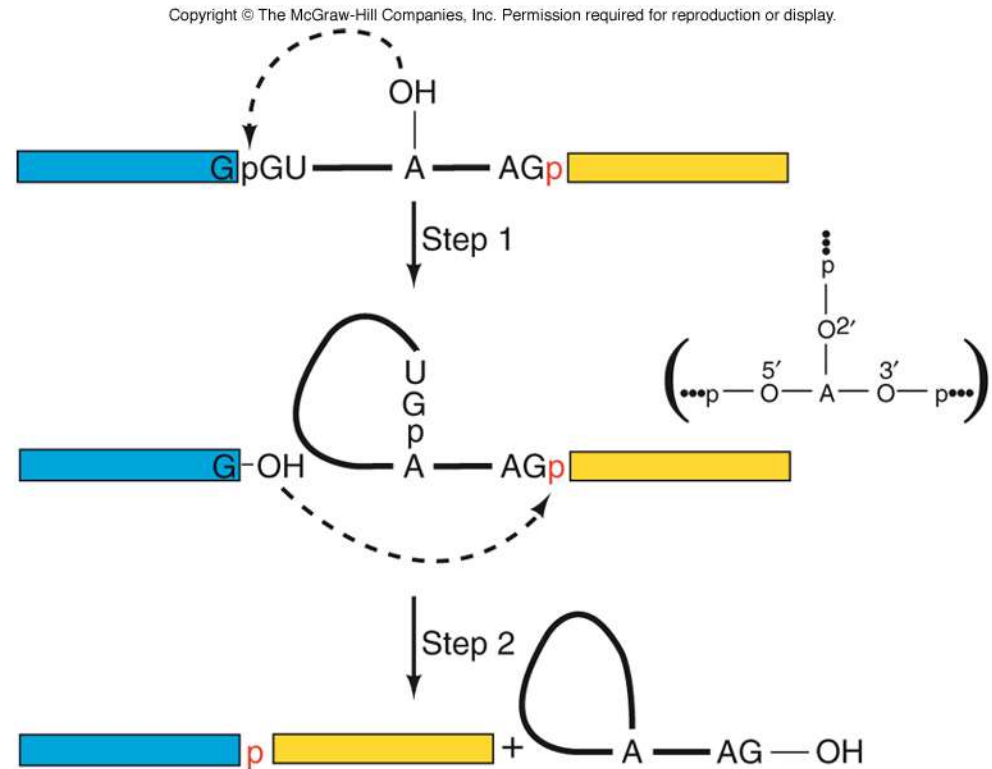
**FIGURE 21.15** Three classes of splicing reactions proceed by two transesterifications. First, a free OH group attacks the exon 1–intron junction. Second, the OH created at the end of exon 1 attacks the intron–exon 2 junction.

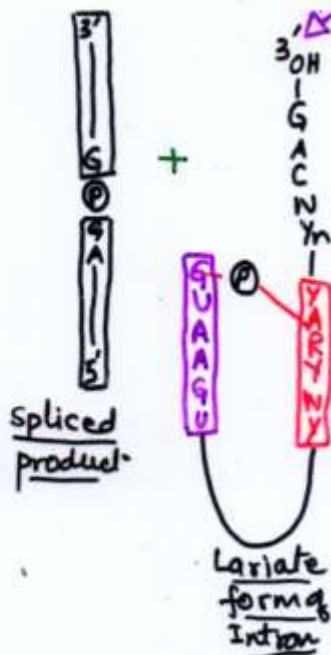
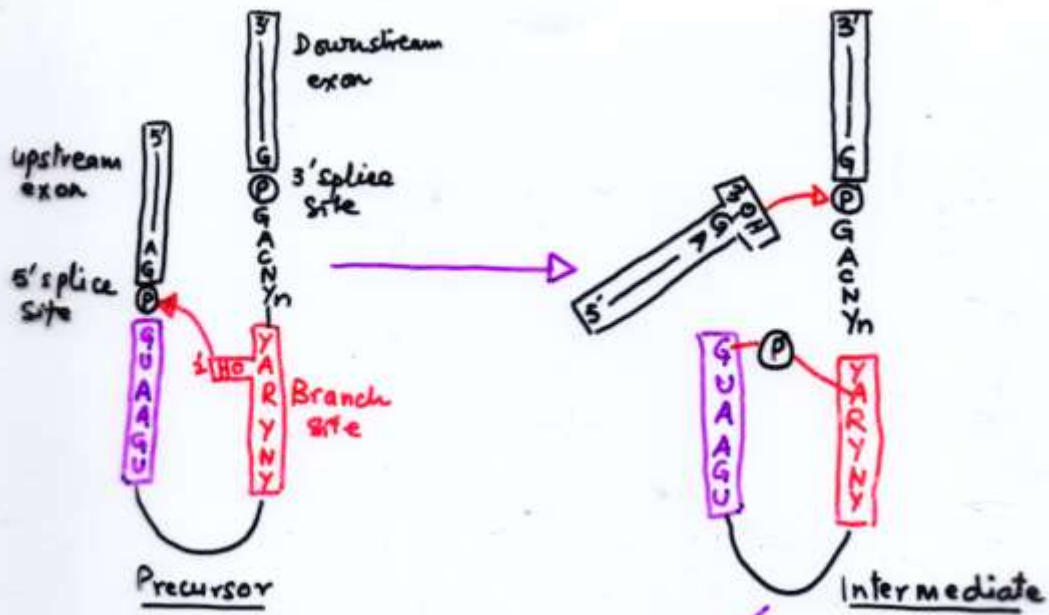
# Splicing Signals: Consensus sequence of the 5' + 3' splice site



# Simplified Mechanism of Splicing

- 2'-OH group of A within intron attacks the phosphodiester bond linking the first exon to the intron
- A lariat is formed due to the GU at the 5' end of the intron forming a phosphodiester bond with the branch point A
- The free 3'OH on exon 1 attacks the phosphodiester bond between the intron and exon 2
- The exons are then linked





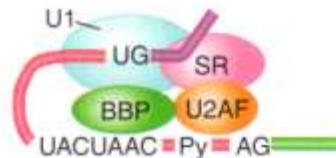
Y = Pyrimidine nucleotide  
 R = Purine nucleotide  
 N = Any nucleotide

Mechanism of Splicing of mRNA precursors in eukaryotic nuclei



## Commitment Complex

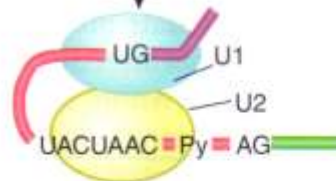
E complex



Formation of commitment  
Complex in which U1 is basepaired  
with the 5' splice site

## Pre-spliceosome

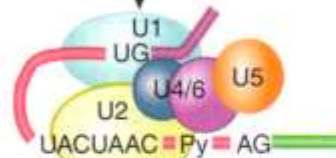
A complex



U2 addition to basepair with the  
branch site in the presence of ATP

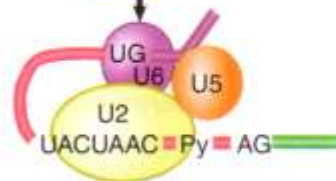
## Spliceosome

B1 complex



Joining of U4.6/U5 tri-snRNPs

B2 complex



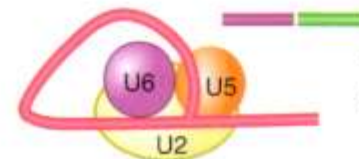
U1 and U4 release  
Formation of the catalytic center  
in which U6 basepairs with the 5' splice site;  
U6 also basepairs with U2; U2 remains  
basepaired with the branch site; U5 interacts  
with both exons through its loop

C1 complex



The first step of transesterification  
5' splice site cleaved, lariat formed

C2 complex



The second step of transesterification  
3' splice site cleaved, exons ligated

**FIGURE 21.12** The splicing reaction proceeds through discrete stages in which spliceosome formation involves the interaction of components that recognize the consensus sequences.



# Commitment, Splice Site Selection and Alternative Splicing

- **snRNPs do not have enough specificity and affinity to bind exclusively and tightly at exon-intron boundaries**
- **Additional splicing factors are needed to help snRNPs bind**
- **Some splicing factors are needed to bridge across introns and exons and so define these RNA elements**

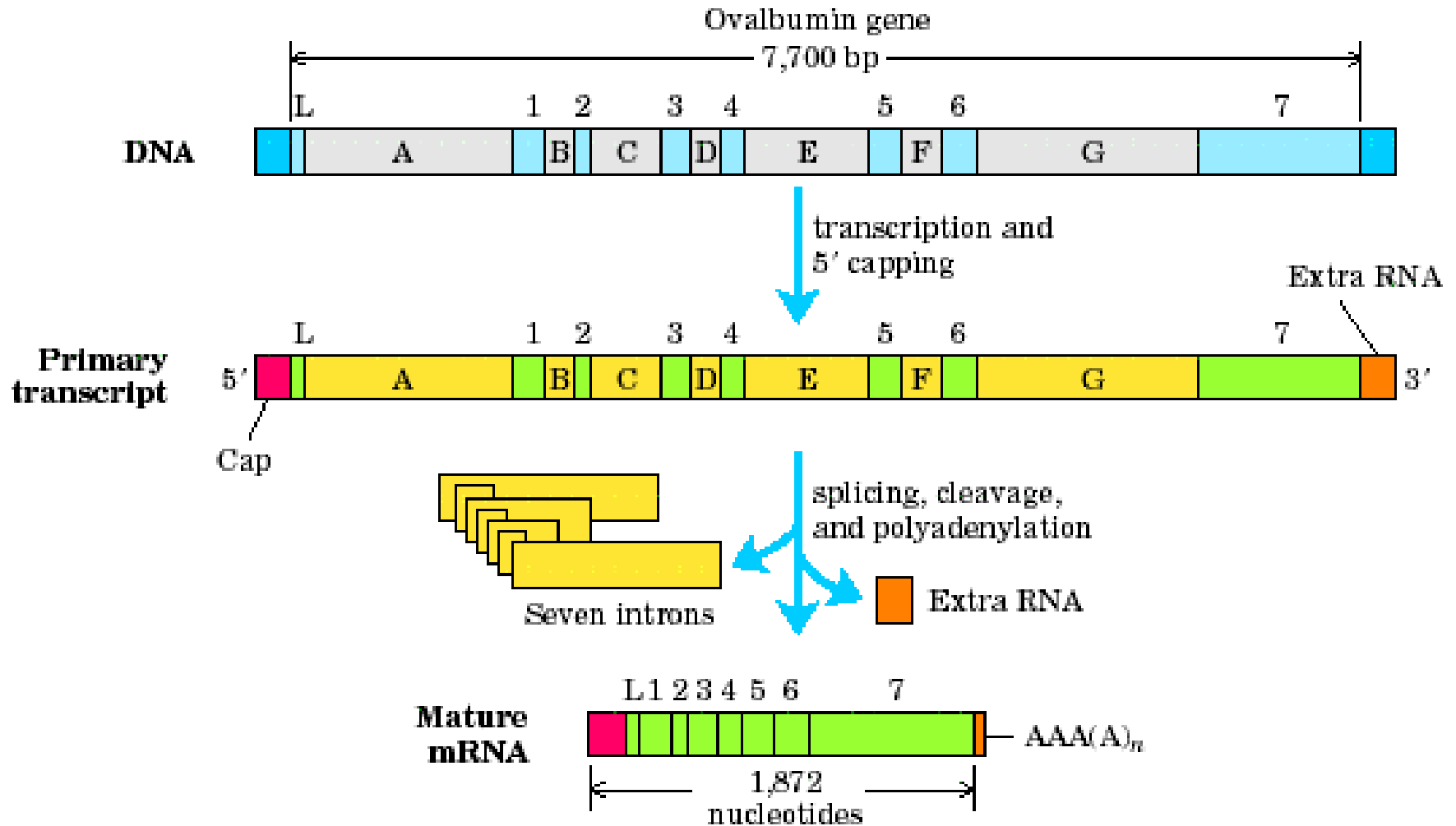
# Exon and Intron Definition

- **The spliceosome can recognize either exons or introns in the splicing commitment process, presumably by assembling splicing factors to bridge across exons or introns**
- **If exons are recognized it is exon definition**
- **If introns are recognized it is intron definition**
- **Splicing in a given organism typically uses either exon definition or intron definition**

## Number of Introns per primary transcript:

<u>Gene</u>	<u># of Introns</u>
Histone & Interferon	0
$\alpha$ -Globin	2
Ig L-Chain	2
Ig H-Chain	4
Yeast mt. cytochrome b	6
Ovomucoid	6
Ovalbumin	7
Ovotransferrin	16
Conalbumin	17
$\alpha$ Collagen	52

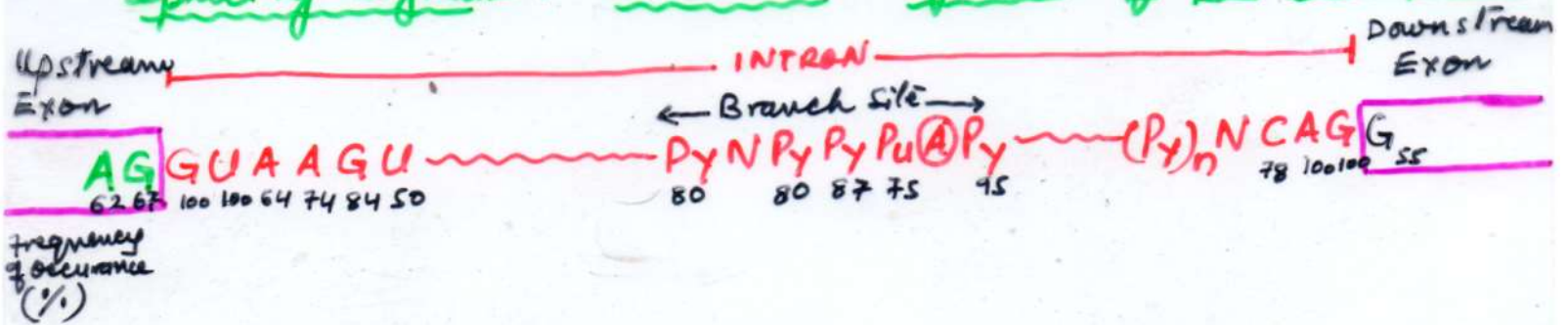
# mRNA Splicing



## Base sequence of intron-exon junctions:

Gene Region	Exon ▼	INTRON	▼ Exon
1. Ovalbumin (Intron-2)	UAAG	<u>GUGAGC</u> ~~~~~	UUACAG GUUG
2. Ovalbumin (Intron-3)	UCAG	<u>GUACAC</u> ~~~~~	AUUCAG UCUG
3. $\beta$ -Globin (Intron-1)	GCAG	<u>GUUGGU</u> ~~~~~	CCUUAG GCUG
4. $\beta$ -Globin (Intron-2)	CAGG	<u>GUGAGU</u> ~~~~~	CCACAG UCUC
5. Ig $\lambda$ , Intron <sub>1</sub>	UCAG	<u>GUCAGC</u> ~~~~~	UUGCAG GGGC
6. SV40 Early Transcript	UAAG	<u>GUAAAU</u> ~~~~~	UUUUAG AUUC

# Splicing Signals: Consensus sequence of the 5' + 3' splice site



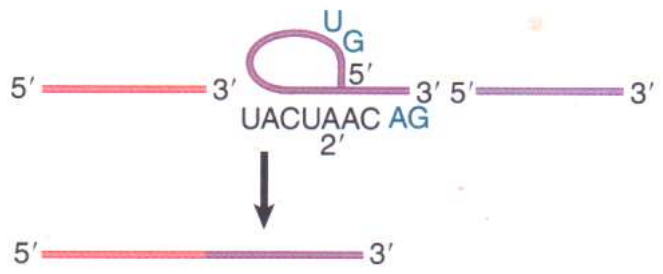


Py<sub>80</sub> N Py<sub>80</sub> Py<sub>87</sub> Pu<sub>75</sub> A Py<sub>95</sub>  
 Animal consensus

Cut at 5' site and form lariat by 5'-2' bond connecting the intron 5'-G to the 2' of A at the branch site



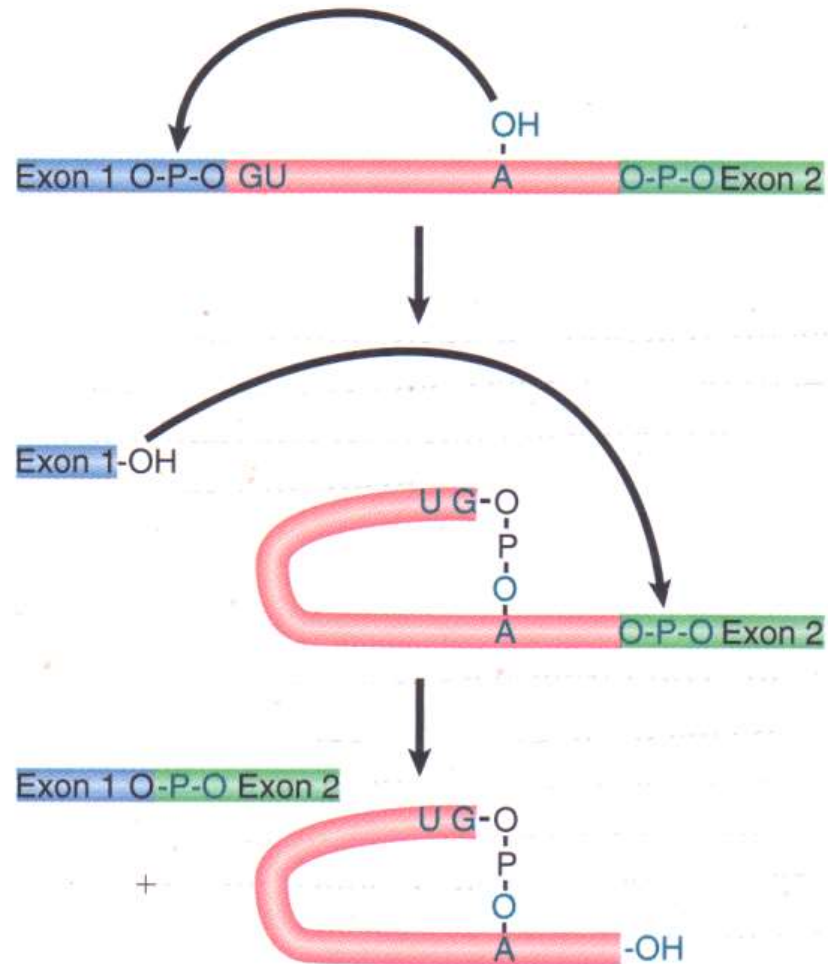
Cut at 3' site and join exons; intron released as lariat



Debranch intron



**FIGURE 21.5** Splicing occurs in two stages. First the 5' exon is cleaved off, and then it is joined to the 3' exon.

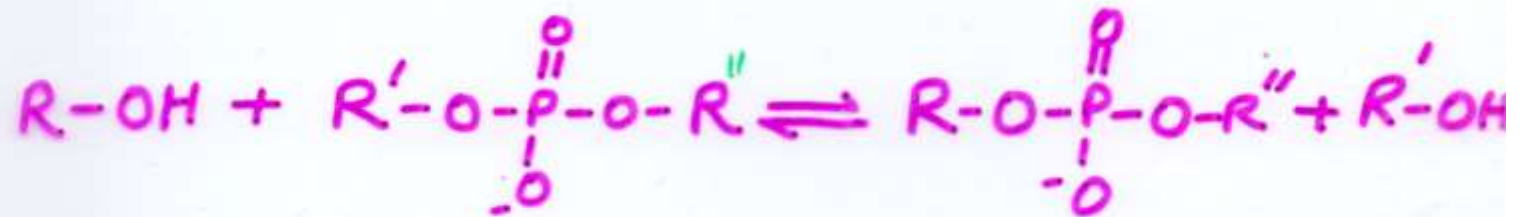


**FIGURE 21.6** Nuclear splicing occurs by two transesterification reactions in which an OH group attacks a phosphodiester bond.



## TRANSESTERIFICATION REACTION

(Transfer of phosphoester)

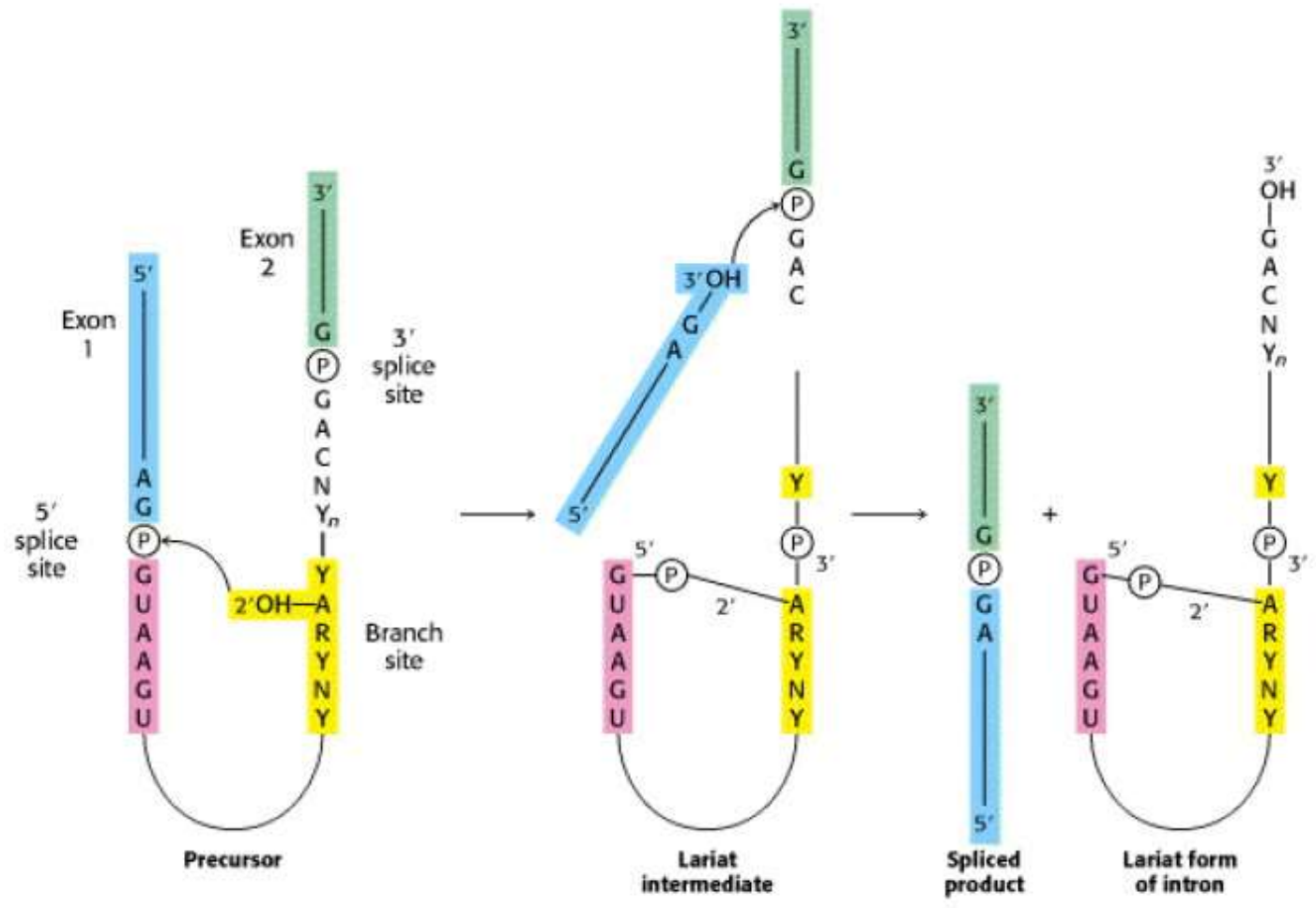


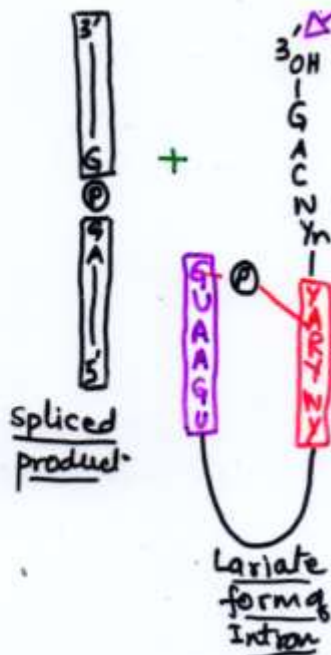
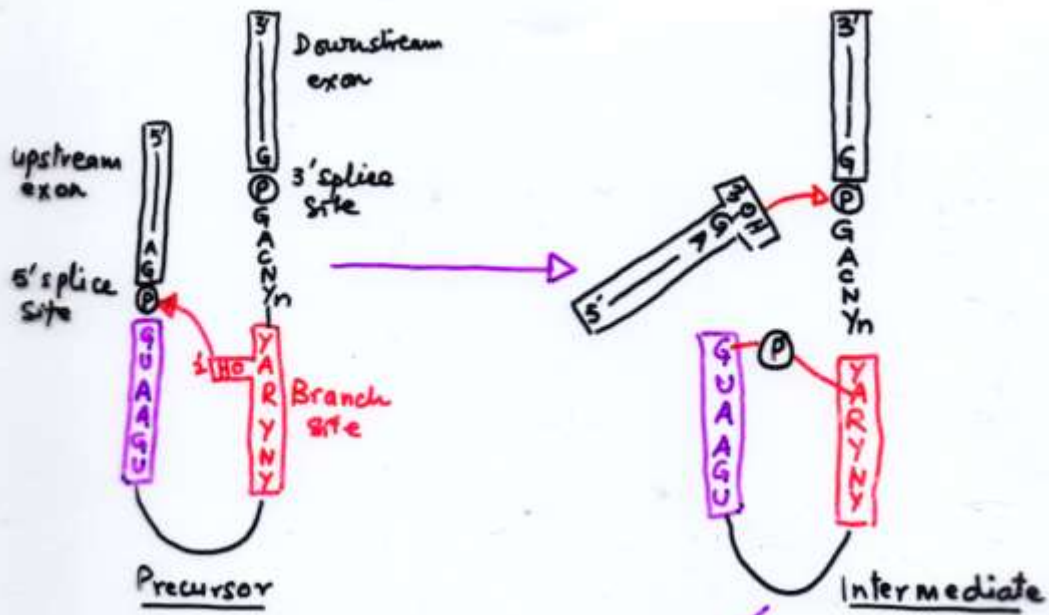
The esters are of phosphate rather than carbon & some of R groups are very long eg -

$R' =$  5' exon, contains ~5,000 nts & has a molecular wt. exceeding 1 million daltons

$R'' =$  1VS + 3' exon

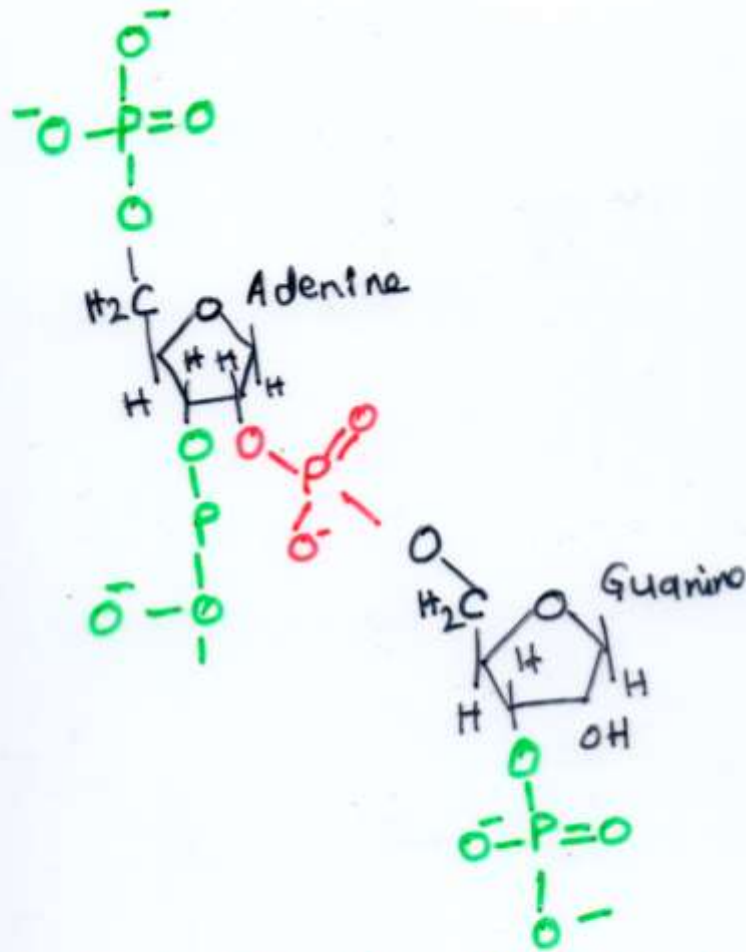
$R' =$  Guanosine or GTP





Y = Pyrimidine nucleotide  
 R = Purine nucleotide  
 N = Any nucleotide

Mechanism of Splicing of mRNA precursors in eukaryotic nuclei



Structure of the Branch Point in the  
lariat intermediate formed during  
splicing

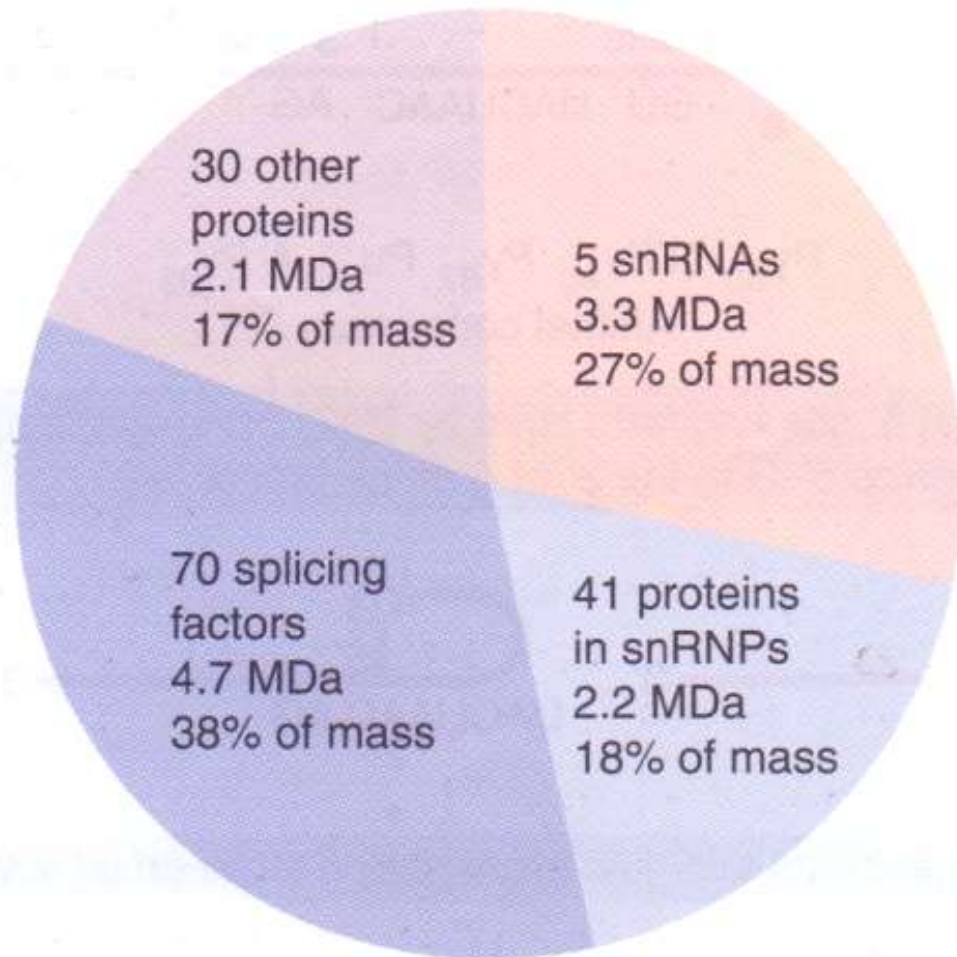
# Role of Branch Site

The role of branch site is to identify the nearest 3' splice site as the target for connection to 5' splice site.

# **Spliceosome Assembly Pathway**

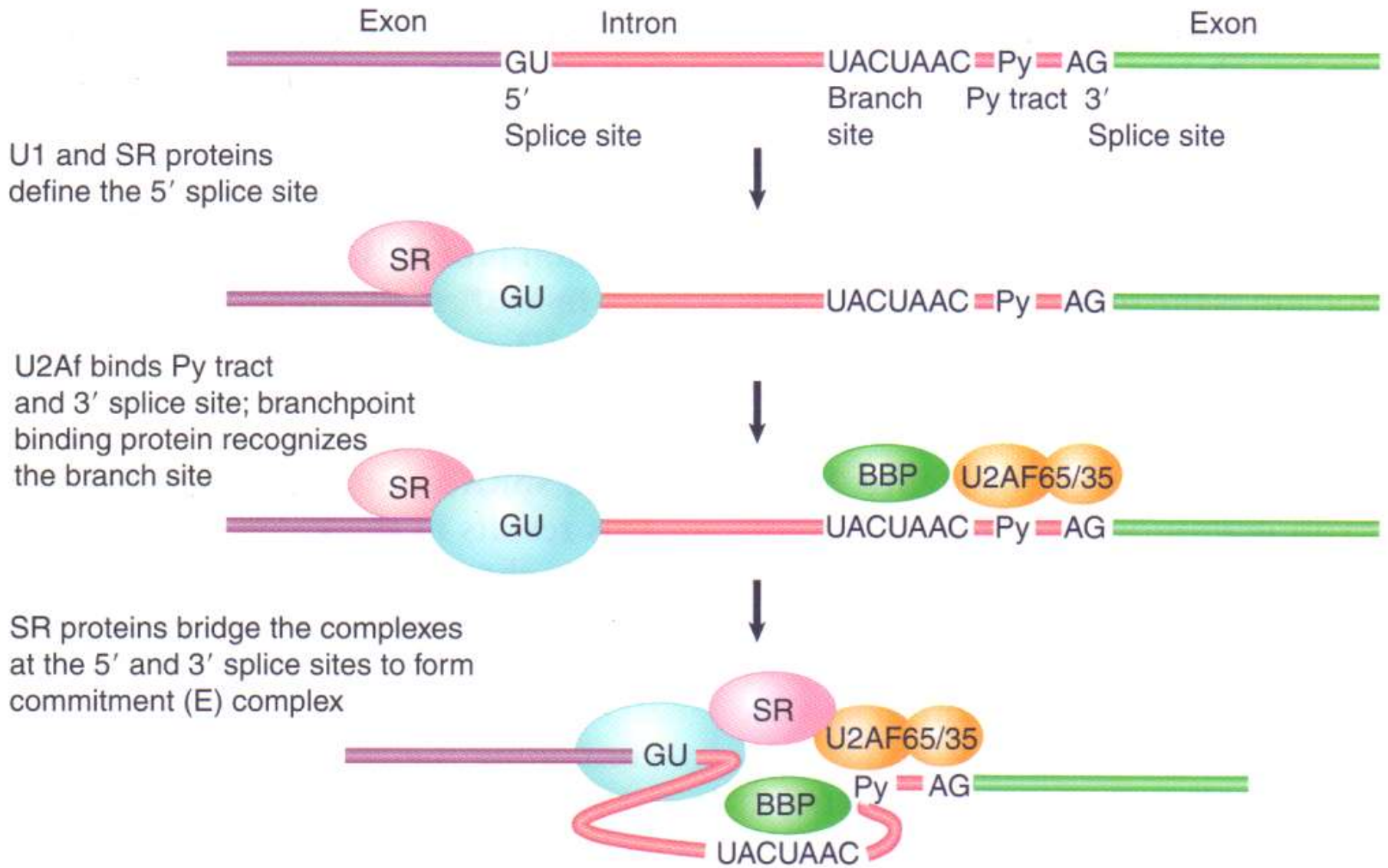
# snRNPs are required for Splicing

- The 5' & 3' splice sites and the branch site are recognized by components of the splicing apparatus. That assemble to form large complex.
- The splicing apparatus contains both proteins and RNAs (in addition to the pre-mRNAs).
- Five snRNAs are involved in splicing reaction e.g., U1, U2, U5, U4 & U6.
- In their natural state, these snRNA exist as ribonucleoprotein particles called snRNPs (U1 snRNP, U2 snRNP, U5 snRNP, U4 snRNP & U6 snRNP)
- SnRNPs associated with splicing signals brings these signals together before any reaction occur.
- These complex assembles sequentially on the pre-mRNA.
- Splicing occurs only after all the components have assembled.



**FIGURE 21.7** The spliceosome is ~12 MDa. Five snRNPs account for almost half of the mass. The remaining proteins include known splicing factors, as well as proteins that are involved in other stages of gene expression.





**FIGURE 21.10** The commitment (E) complex forms by the successive addition of U1 snRNP to the 5' splice site, U2AF to the pyrimidine tract/3' splice site, and the bridging protein SF1/BBP.

### Intron definition



Interactions across the intron



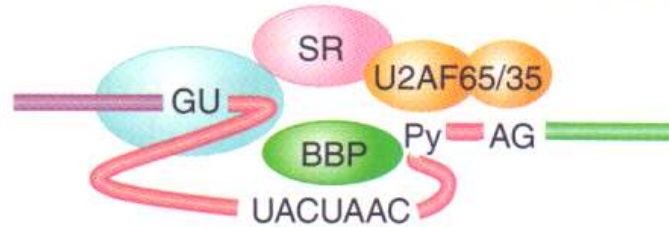
### Exon definition



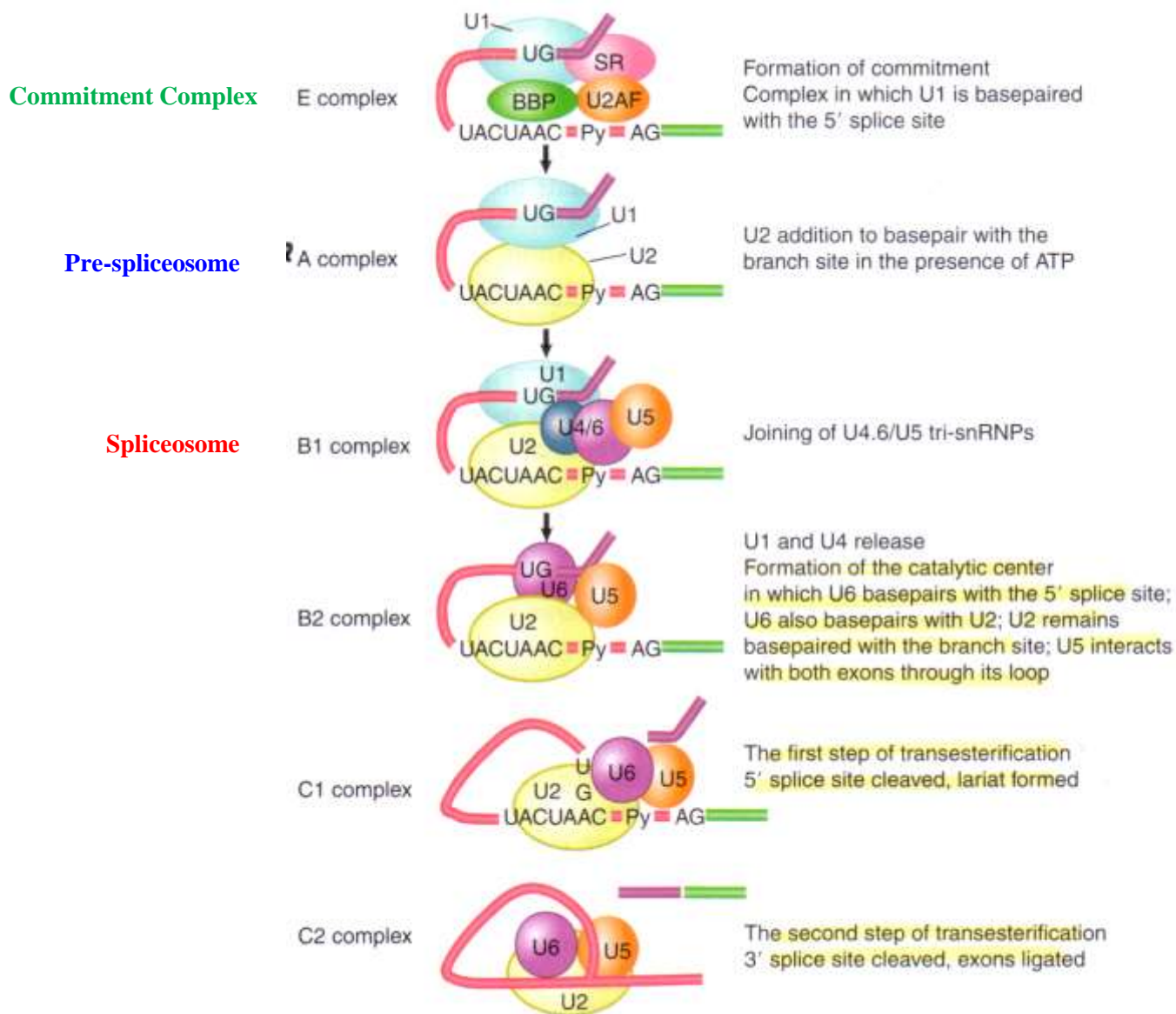
Interactions across the exon



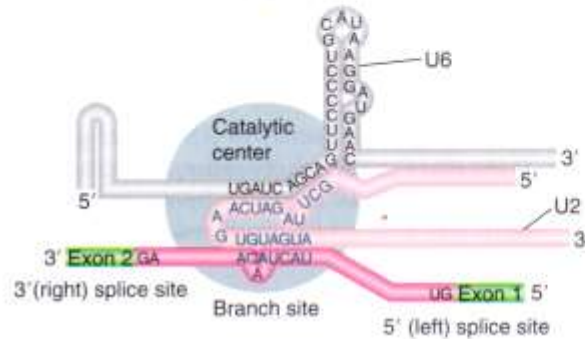
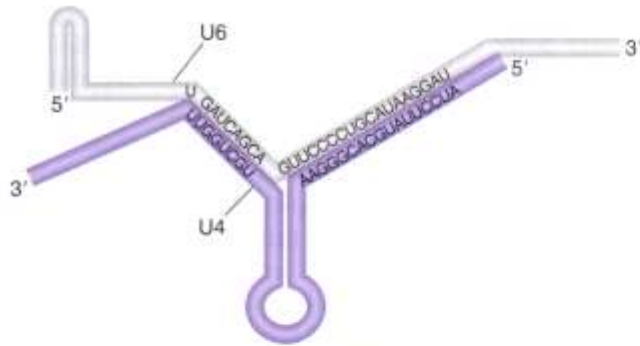
Switch from interactions across the exon to those across the intron



**FIGURE 21.11** There are two routes for initial recognition of 5' and 3' splice sites by either intron definition or exon definition.



**FIGURE 21.12** The splicing reaction proceeds through discrete stages in which spliceosome formation involves the interaction of components that recognize the consensus sequences.

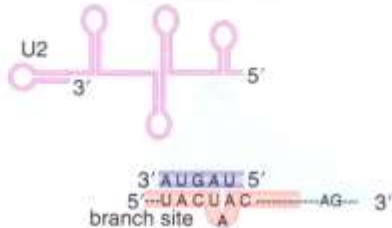


**FIGURE 21.13** U6-U4 pairing is incompatible with U6-U2 pairing. When U6 joins the spliceosome it is paired with U4. Release of U4 allows a conformational change in U6; one part of the released sequence forms a hairpin (gray), and the other part (pink) pairs with U2. An adjacent region of U2 is already paired with the branch site, which brings U6 into juxtaposition with the branch. Note that the substrate RNA is reversed from the usual orientation and is shown 3' to 5'.

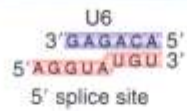
U1 pairs with the 5' splice site



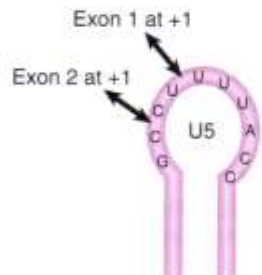
U2 pairs with the branch site



U6 pairs with the 5' splice site



U5 is close to both exons



**FIGURE 21.14** Splicing utilizes a series of base pairing reactions between snRNAs and splice sites.